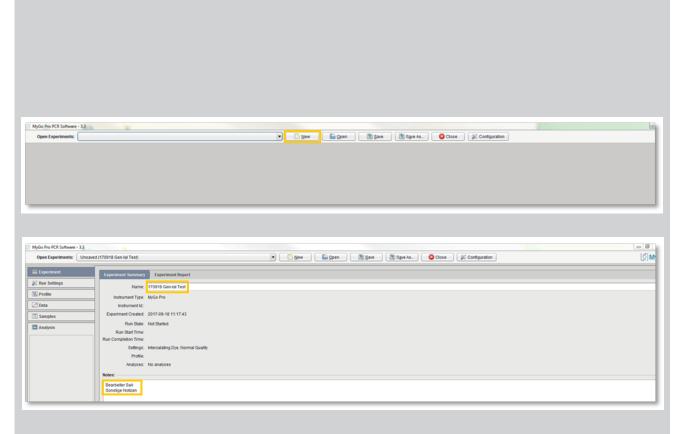




Programming set-up for MyGo Pro for CONGEN SureFood®/SureFast® kits (2-3 plex kits)*

*Please note: 4plex SureFood®/SureFast® kits may not be used on this thermocycler





Step 1:

Open the MyGo Pro Software, click on "New" to set-up the required detection format template.

Enter the name of the data file in "Notes"

Optional enter the username and the Lot No. of the particular kit in the "Notes" window.

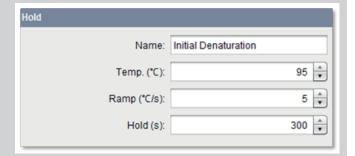
MyGo Pro PCR Software - 3.3 Open Experiments: Unsaved (170918 Gen-lal Test) New ☐ Open ☐ Save ☐ Save As... ☐ Close ☐ Configuration Experiment Please select the brightest dye present in any well. If you are unsure, use the "intercalating Dyes" option. V. Profile Intercalating Dyes (SYBR Green I, ResoLight) Data Hydrolysis Probes (FAM, VIC, HEX, Yellow 555, Red 610, TexasRed, Cy5) Samples Other Dyes Analysis The instrument can increase signal quality by acquiring for longer on each cycle of amplification, after the hold is complete. Please select desired signal quality - if you are unsure, use the 'Normal Quality' option. The instrument can automatically find the best settings to use during melting. If Optimize Meit Acquisitions is selected, the acquisition time will depend on dije intensity. If the option is not selected, the settings above will be used throughout the run. Optimised integration time will be displayed Optimize Melt Acquisitions Use Advanced Settings Display Dye Calibrations MyGo Pro PCR Software - 3.3 Dpen Open Open Experiments: Unsaved (170918 Gen-lal Test) New New Save Experiment 0.5 Integration Time (s): **II.** Profile 10 🗘 Acquisitions per Cycle: **Data** The instrument can automatically find the best settings to use during melting. If Optimize Melt Acquisitions is selected, the acquisition time will depend on dye intensity. If the option is not selected, the settings above will be used throughout the run. Optimised integration time will be displayed Samples in the log. Optimize Melt Acquisitions Analysis

Step 2:

Navigate to "Run Settings" and choose "Hydrolysis Probes" and "Normal Quality".

Choose "Use Advanced Settings" and set "Integration Time" to 0.5 und "Acquisitions per Cycle" to 10.

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Step 3:

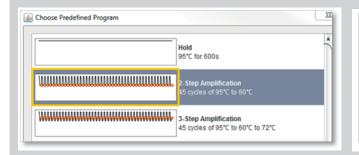
Navigate to "Profile".

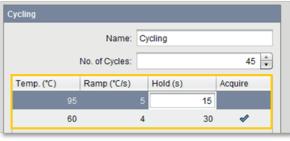
Enter the run protocol according to the SureFast®/SureFood® assay.

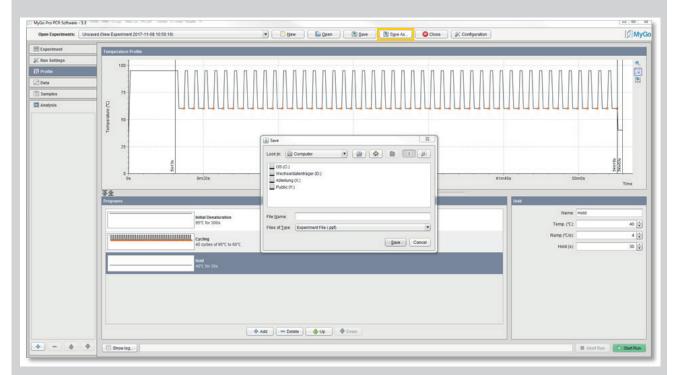
Start with "Add" and choose the "Hold" button for the first step of PCR Cycling "Initial Denaturation"

Following the run protocol thus adding new cycling steps through "Add".

	Blockcycler / LightCycler® 480*
Initial Denaturation (HOLD)	5 min, 95°C
Cycles	45
Denaturation	15 sec, 95°C
Annealing/Extension (CYCLE)	30 sec, 60°C







Step 4:

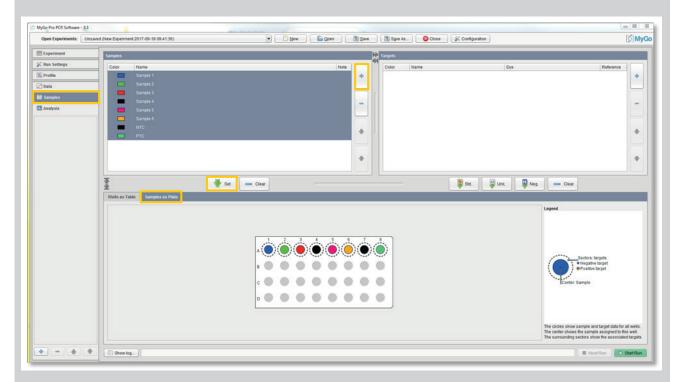
Click "Add" and choose "2-Step Amplification" for defining the "cycling" according to the setup of the real-time PCR manual.

The measurement of the fluorescence signal should always take place at the end of the annealing/ extension step.

For example: SureFood® Animal ID horse & donkey 3plex S6119 select for the No. of cycles 35; set the denaturation time to 15 sec and the temperature to 60 °C.

Optional add a "Hold" step for 30 sec at 40 °C to cool the tubes at the end of the setup.

Finally save the setup on your computer.

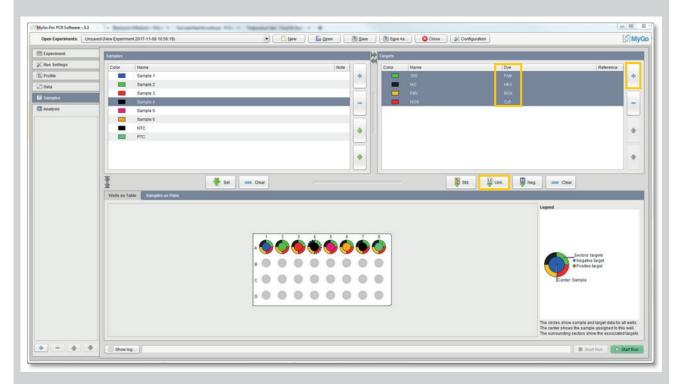


Step 5:

Navigate to "Samples" and select "+" on the left side of the window for every sample including controls (e.g. PTC; NTC; EC). Name your sample subset.

Choose "Samples as Plate" and mark the samples together with the particular position in the cycler and source the plate with "Set".

Every sample should be highlighted in the plate section as a colored circle.



Step 6:

Navigate to the right window and choose "+" to enter the unique detection format. For 3plex assays as SureFood®Animal ID S6119 select the four Dyes: FAM, HEX, ROX in the drop-down menu.

Name the detection channels according to the targets of the SureFood®/SureFast® assay.

Optional define the Color of the dyes green for FAM, black for IAC, orange for ROX.

Mark all targets and choose "Unk".

Every colored circle should be sorrounded by a four-color border.

Save the experiment and start the run.



R-Biopharm Contacts:

Information:

Phone: +49 (0) 61 51 - 81 02-0
Fax: +49 (0) 61 51 - 81 02-40
Fax: info@n his a harmen de

E-mail: <u>info@r-biopharm.de</u>

Orders:

Phone: +49 (0) 61 51 - 81 02-0 Fax: +49 (0) 61 51 - 81 02-20 E-mail: <u>orders@r-biopharm.de</u>

www.r-biopharm.com